

## Research Article

### Prevalence of Gastrointestinal Parasites of Stray Dogs in Suleja Metropolis, Niger State, Nigeria

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## ABSTRACT

Free-roaming dogs in the environment (stray dogs) have been reported to be a reservoir of various gastrointestinal (GIT) parasites of significant public health importance worldwide. This study aims to determine the prevalence of GIT parasites in the stray dog population in Sabo Gwazunu, Suleja and within Suleja Township, of Niger State, Nigeria. Sixty fecal samples were randomly collected from stray dogs in each study area (community) between February and March 2024. The Sheather's sugar floatation technique was used to examine various faecal samples, and the prevalence was calculated as the percentage of infected samples over the total number of samples examined. Overall, a total of thirteen GIT parasites belonging to Phylum Protozoa 3.33% (4/120), Platyhelminthes 3.33% (4/120), and Nematoda 4.16% (5/120), were found. A significant difference was observed in GIT prevalence between the study areas ( $P < 0.05$ ). GIT parasites found in stray dogs within Suleja township comprise *Toxocara canis*, *Eimeria* sp, *Strongyloides* sp, *Alaria* sp, *Cryptosporidium* sp, *Entamoeba* sp, *Ancylostoma* sp, *Dipylidium* sp, *Schistosoma* sp, and *Taenia* sp. However, in Sabo Gwazunu, nine GIT parasites were found in stray dogs. These include *Eimeria* sp, *T. canis*, *Trichuris* sp, *Physaloptera* sp, *Strongyloides* sp, *Dipylidium* sp, *Schistosoma* sp, *Alaria* sp, and *Taenia* sp. Moreover, the prevalence of GIT in stray dogs in both communities was estimated at 91% (91/120). Given the public health and zoonotic significance of some gastrointestinal parasites observed in this study, it is essential to increase awareness about the need to control the stray dog population in the study areas.

**Keywords:** Gastrointestinal parasites; Stray dogs; Sabo Gwazunu; Suleja

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## INTRODUCTION

Gastrointestinal parasites (GIT) pose a significant health concern in dogs and especially in stray dogs (Ugbomoiko *et al.* 2008; Lyons *et al.* 2022; Kirman *et al.* 2022). These parasites can cause a range of health problems and are a significant issue in many areas where stray dogs are prevalent (Lyons *et al.* 2022). In Nigeria, the prevalence

of gastrointestinal parasites in stray dogs is a pressing concern due to factors like the tropical climate which enhance transmission and the large stray dog population in the country. The occurrence of these parasites in dogs can vary depending on regional and environmental conditions (Odeniran *et al.*, 2013; Ezema *et al.*, 2019). For example, stray dogs in urban areas may

exhibit different parasite profiles than those in rural regions, with higher prevalence often found in areas lacking proper sanitation and veterinary care access. Other contributing factors to the disparity include population density and human activities (Odeniran *et al.*, 2013).

In Nigeria, common gastrointestinal parasites affecting stray dogs include roundworms (*Toxocara canis*), hookworms, tapeworms, Echinococcus (hydatid tapeworm), whipworms, Giardia, and coccidia. Many of these parasites pose significant economic and public health concerns. (Ugbomoiko *et al.* 2008; Abbas *et al.*, 2023). Hookworms are blood-sucking parasites that attach to a dog's intestinal lining and can lead to anemia and other health issues. Dogs may become infected through contaminated soil, skin penetration, ingestion of larvae from the environment, or through the milk of infected mothers, all of which can result in harmful effects (Bhattarai *et al.*, 2020 Abbas *et al.*, 2023).

The harmful effects of gastrointestinal parasites usually extend beyond their hosts (dogs), as they can be major contributors to zoonotic diseases (Del Brutto *et al.*, 2018; Bhattarai *et al.*, 2020). Zoonotic infections can be acquired through direct contact with animals, contaminated water, contaminated food, and indirect contact with animal secretions and excretions. Besides hookworms, *T. canis* is another GIT parasite that causes various clinical issues, such as visceral larva migrans and ocular larva migrans. *T. canis* infection has been associated with cognitive impairments and developmental delays, particularly in young children. Research suggests that *T. canis* antibodies in children are linked to lower cognitive scores, poor attention span, and learning difficulties (Bhattarai *et al.*, 2020). Furthermore, individuals infected with *T. canis* may develop allergic reactions due to sensitization to the parasite's antigens, leading to symptoms such as allergic asthma, wheezing, skin rashes, or eosinophilia. (Rubinsky-Elefant *et al.*, 2010; Mizgajska-Wiktor *et al.*, 2017). Usually, the impact of *T. canis* on public health extends beyond direct health impacts. The expenses related to diagnosing and treating *T. canis* infections, especially in severe cases, can place a substantial economic strain on healthcare systems and affected individuals. (Rubinsky-Elefant *et al.*, 2010; Omonijo and Mukaratirwa, 2023). Human infection occurs when the infectious stage of *T. canis* is accidentally ingested from a contaminated environment, through consumption of contaminated food or water, or by eating encapsulated larvae in raw or undercooked tissues of paratenic hosts. (Rubinsky-Elefant *et al.*, 2010; Omonijo and Mukaratirwa, 2023).

Elderly individuals, young children, and those with weakened immune systems are at the highest risk of

contracting zoonotic infections (Del Brutto *et al.*, 2018; Bhattarai *et al.*, 2020). Given the economic and public health concerns associated with gastrointestinal parasites in dogs, research should focus on the prevalence of these parasites in stray dogs to determine the environmental burden of infection. The present study therefore aimed to establish baseline data on the prevalence of gastrointestinal parasites affecting stray dogs in the Suleja metropolis.

## **MATERIALS AND METHODS**

### **Study Areas**

The study was conducted in Sabo (new) Gwazunu, Suleja and within Suleja Township, Suleja Local Government Area of Niger State, Nigeria (Figure 1). Suleja Township study area is found at latitude 9°10'50.12" N and longitude 7° 10' 45.80" E. The density of vegetation cover in the area has been greatly reduced as a result of urbanization. New Gwazunu is a resettlement area south of Suleja township (Latitude: 9°08'13.18"N; Longitude: 7° 11' 46.79"E). Typical vegetation of lush grassland with many trees, shrubs, and bushes can be seen (Adesoye *et al.*, 2024). Generally, most people in Suleja metropolis rear animals, including dogs with majority engaging in agricultural activities.

### **Sample Collection**

Sixty (60) faecal samples were randomly collected from stray dogs on the streets of each community immediately after defecation, between February and March 2024. The faecal were put into plastic universal bottles, properly labeled, and transported to the laboratory of Centre for Tuberculosis and other Neglected Tropical Diseases, Suleja, for analysis. The samples were refrigerated at 4°C and held until the following day, when they were processed for ova and oocyst detection.

### **Parasitology Procedure**

Faecal samples were processed individually and examined microscopically using Sheather's sugar technique (Duncan *et al.*, 2020). 1 g of faecal was added to 15 ml of sheather's sugar solution in a test tube. The content was thoroughly mixed and strained, collected into another test tube, and covered with a slip. The filtrate was left to stand for 15 – 30 mins. The cover slip was then transferred to a microscopic slide to be examined under a light microscope. The parasite samples were identified using the standard key procedure (Cimino *et al.* 2015).

### **Data Analysis**

Data analysis was conducted using IBM-SPSS version 25.0. Chi-square tests were applied to assess the association between communities and parasite prevalence, with statistical significance set at  $P < 0.05$ .

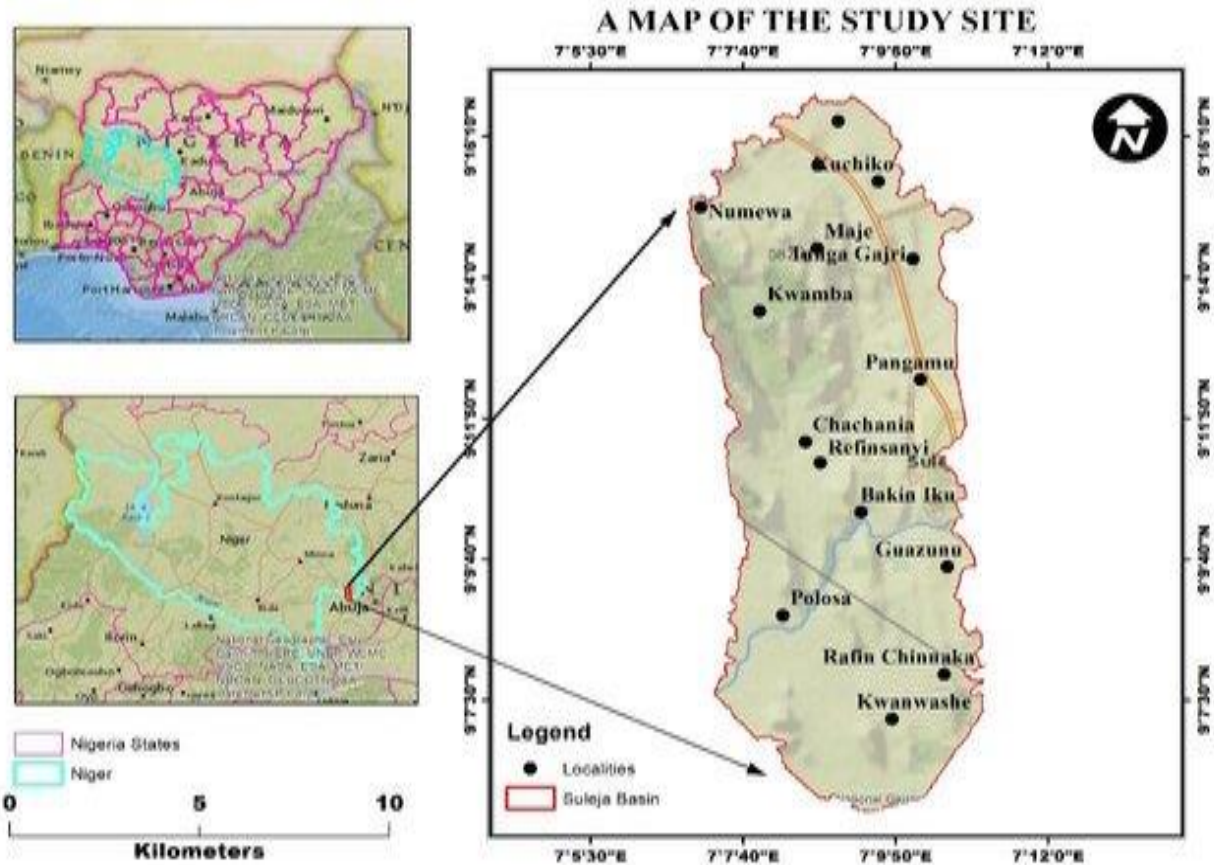


Figure 1. Map of Nigeria and Suleja Metropolis

**RESULTS**

**Prevalence of Gastrointestinal Parasites from Stray Dogs in Suleja Township and Sabo Gwazunu, Niger State**

A total of thirteen species gastrointestinal parasites belonging to Phylum Protozoa (*Cryptosporidium* sp and *Eimeria* sp., *Entamoeba* sp and *Hartmannella* sp), Phylum Nematoda (*T. canis*, *Trichuris* sp, *Physaloptera* sp, *Strongyloides* sp, *Ancylostoma* sp), and Phylum Platyhelminthes (*Alaria* sp, *Dipylidium* sp, *Schistosoma* sp, and *Taenia* sp) were found in stray dogs in both Suleja township and Sabo Gwazunu. In Suleja Township, eleven species GIT parasites were found, and these include *Hartmannella* sp, *Eimeria* sp, *T. canis*, *Strongyloides* sp, *Alaria* sp, *Cryptosporidium* sp, *Entamoeba* sp, *Ancylostoma* sp, *Dipylidium* sp, *Schistosoma* sp, and *Taenia* sp. However, in Sabo Gwazunu, only nine species GIT parasites were found. These include *Eimeria* sp, *T. canis*, *Trichuris* sp, *Physaloptera* sp, *Strongyloides* sp, *Dipylidium* sp, *Schistosoma* sp, *Hartmannella* sp, and *Taenia* sp. (Table 1). The prevalence of GIT parasites between the two

study sites showed a significant difference ( $P<0.05$ ). Across the two study communities, *T. canis* was observed to have the highest prevalence of 46.66% (28/100), followed by *Dipylidium* sp, 30.00% (18/120), *Taenia* sp, Overall, the prevalence of GIT parasites in both communities was estimated at 75.83% (91/120).

**Prevalence of parasitic infection in stray dogs in Suleja Township**

Table 2 shows the infection status of stray dogs in Suleja Township. Out of the sixty faecal samples of stray dogs that were examined, 11 (18.33%) are without GIT parasitic infection, 37 (72.0%) have one GIT parasite, and just three of the samples have double GIT parasitic infection, namely *Taenia* sp + *Toxocara canis*, *Taenia* sp + *Ancylostoma* sp and *Taenia* sp + *Strongyloides* sp.

**Prevalence of parasitic infection in stray dogs in Sabo Gwazunu**

Ten (16.66%) out of the entire 60 fecal samples collected in Sabo Gwazunu were uninfected with GIT parasites. Single infection constituted 49 (81.66%), while double infection consisted of two samples (3.33%): *Schistosoma* sp + *T. canis* and *Taenia* sp + *T. canis* (Table 3).

**Table 1. Prevalence of gastrointestinal parasites in Suleja Metropolis**

| Parasite species          | Communities            |                     |                        |
|---------------------------|------------------------|---------------------|------------------------|
|                           | Suleja Township (N=60) | Sabo Gwazunu (N=60) | Overall prevalence (%) |
| <i>Eimeria</i> sp         | 2 (3.33)               | 7 (11.66)           | 9 (15.00)              |
| <i>Hartmannella</i> sp    | 1 (1.66)               | 1 (1.66)            | 2 (3.33)               |
| <i>Toxocara canis</i>     | 6 (10.00)              | 22 (36.66)          | 28 (46.66)             |
| <i>Trichuris</i> sp       | 0 (0.00)               | 1 (1.66)            | 1 (1.66)               |
| <i>Physaloptera</i> sp    | 0 (0.00)               | 1 (1.66)            | 1 (1.66)               |
| <i>Strongyloides</i> sp   | 4 (6.66)               | 3 (5.00)            | 7 (11.66)              |
| <i>Alaria</i> sp          | 1 (1.66)               | 0 (0.00)            | 1 (1.66)               |
| <i>Cryptosporidium</i> sp | 2 (3.33)               | 0 (0.00)            | 2 (3.33)               |
| <i>Entamoeba</i> sp       | 1 (1.66)               | 0 (0.00)            | 1 (1.66)               |
| <i>Ancylostoma</i> sp     | 4 (6.66)               | 0 (0.00)            | 4 (6.66)               |
| <i>Dipylidium</i> sp      | 12 (20.00)             | 6 (10.0)            | 18 (30.00)             |
| <i>Schistosoma</i> sp     | 1 (1.66)               | 3 (5.00)            | 4 (6.66)               |
| <i>Taenia</i> sp          | 4 (6.66)               | 9 (15.00)           | 13 (21.66)             |
| <b>Total</b>              | <b>38 (63.30)</b>      | <b>53 (88.00)</b>   | <b>91 (75.83)</b>      |

**Table 2. Infection Prevalence among stray dogs with gastrointestinal parasites in Suleja Township, Niger State**

| Infection status                           | Prevalence (%) |
|--|----------------|
| No infection                               | 11 (18.33%)    |
| Single infection                           | 37 (72.0%)     |
| <i>Alaria</i>                              | 1 (1.66)       |
| <i>Ancylostoma</i> sp                      | 3 (5.00)       |
| <i>Cryptosporidium</i> sp                  | 2 (3.33)       |
| <i>Dipylidium</i> sp                       | 12 (20.00)     |
| <i>Hartmannella</i> sp                     | 1 (1.66)       |
| <i>Eimeria</i> sp                          | 2 (3.33)       |
| <i>Entamoeba</i> sp                        | 1 (1.66)       |
| <i>Schistosoma</i> sp                      | 1 (1.66)       |
| <i>Strongyloides</i> sp                    | 3 (5.00)       |
| <i>Taenia</i> sp                           | 1 (1.66)       |
| <i>Toxocara canis</i>                      | 5 (8.33)       |
| Multiple Infection                         | 3 (5.00)       |
| <i>Taenia</i> sp + <i>Toxocara canis</i>   | 1 (1.66)       |
| <i>Taenia</i> sp + <i>Ancylostoma</i> sp   | 1 (1.66)       |
| <i>Taenia</i> sp + <i>Strongyloides</i> sp | 1 (1.66)       |

**Table 3. Infection Prevalence among stray dogs with gastrointestinal parasites in Sabo Gwazunu, Niger State**

| Infection status        | Number of samples (%) |
|-------------------------|-----------------------|
| No infection            | 10 (16.66)            |
| Single infection        | 49 (81.66)            |
| <i>Dipylidium</i> sp    | 6 (10.00)             |
| <i>Hartmannella</i> sp  | 1 (1.66)              |
| <i>Eimeria</i> sp       | 7 (11.66)             |
| <i>Physaloptera</i> sp  | 1 (1.66)              |
| <i>Schistosoma</i> sp   | 2 (3.33)              |
| <i>Strongyloides</i> sp | 3 (5.00)              |
| <i>Taenia</i> sp        | 8 (13.33)             |
| <i>Toxocara canis</i>   | 20 (33.33)            |

|                                  |          |
|----------------------------------|----------|
| <i>Trichuris sp</i>              | 1 (1.66) |
| Multiple infections              | 2 (3.33) |
| <i>Schistosoma sp + T. canis</i> | 1 (1.66) |
| <i>Taenia sp + T. canis</i>      | 1 (1.66) |

## DISCUSSION

Studying the prevalence of gastrointestinal parasite in dogs is crucial for protecting both human and animal health, and it supports the development of policies to minimize the spread of zoonotic diseases. Such research works has significant public health implications, especially because many of these parasites can be transmitted from animals to their immediate human population (Bhattarai, 2020; Kamani *et al.*, 2021).

There has been a number of reports on prevalence of gastrointestinal parasites in Africa (Sulieyman *et al.* 2020) and most especially in Nigeria (Ayinmode *et al.* 2016; Moro and Abah, 2019; Kamani *et al.*, 2021). However, there is little or no information on the prevalence of GIT parasites in Niger State. Findings from this study show a prevalence of 75.83% of GIT parasites in stray dogs in Suleja metropolis. This is higher than the prevalence of 39.8% (377/948) reported in a nationwide survey in Nigeria (Kamani *et al.*, 2021). The disparity observed in the study may be as a result of the number of dogs higher in Nigeria as a whole country compared to a locality in Niger State. The higher the number hosts, the greater the possibility of parasite prevalence in an environment (Wu *et al.* 2019). There is also such difference in the prevalence of parasite in Suleja metropolis and 43.3% (88/203) in Ibadan, Oyo State (Ayinmode *et al.* 2016), 65% (260/400) in Abua, Rivers State (Moro and Abah, 2019), 68.4% (271/396) in Kwara State (Ugbomoiko *et al.* 2008), 73.3% (293/400) in Makurdi metropolis, Benue State (Matthew *et al.* 2016), 77.9% (366/470) in Northeastern Nigeria (Magaji *et al.* 2012), and 78.85% (41/52) in Kebbi State (Jajere *et al.* 2022).

Results from the present study showed higher 88.00% (53/120) parasitic prevalence in Sabo Gwazunu compared to 63.30% (38/120) in Suleja township. Bushes and native vegetation can be found in abundance at Sabo Gwazunu compare to the later location. This may have accounted for higher parasite prevalence in Sabo Gwazunu. There are reports associating poor hygiene and sanitation with higher prevalence of endoparasite in an environment (Archer *et al.* 2017; Issa *et al.* 2022). Stafford *et al.*, (2020) findings buttress the fact that neater climes do have lower prevalence of parasite as compared to dirty ones. One could deduce lower prevalence of 11.9% (43/360) in their report is the fact that United States has not only limited stray dogs but also significant neater

environment. Some other countries of the world have a lower reported prevalence as compared to the result of the present study, as such, 22.66% (80/353) in Cuiabá, Brazil (Souza *et al.* 2023); Turkey (Ünal *et al.* 2022); 46.25% in Zakho city, Iraq (Issa *et al.* 2022) and 59.50% (238/400) in Suryabinayak, Nepal (Sukupayo and Tamang, 2023); 63.94% (172/269) in Kurdistan region. This observation is consistent with a report from an earlier study, which showed that the prevalence of gastrointestinal parasites in dogs may vary by region and environmental conditions (Archer *et al.* 2017; Ezema *et al.* 2019).

Furthermore, this study showed that among the GIT parasites found, *Toxocara canis* has the highest prevalence, 56.0% (28/120). This is consistent with report from Kebede (2019) that reported a high prevalence of *T. canis* in the dog population. However, some studies reported lower prevalence (Sowemimo, 2009, Akeredolu and Sowemimo, 2014), while another reported no *T. canis* prevalence (Sulieyman *et al.* 2020). *T. canis* is a zoonotic nematode responsible for larva migrans syndrome (LMS), visceral larva migrans (VLM), ocular larva migrans (OLM), neural larva migrans (NLM), and covert infection in humans (Phoosangwalthong *et al.* 2022). Humans can become infected with these parasites by consuming contaminated eggs from soil or water, through unwashed hands, raw vegetables, or by ingesting larvae present in undercooked or raw infected organ or muscle tissues of other paratenic hosts (Phoosangwalthong *et al.* 2022). This study highlights the importance of preventing stray dogs from accessing the environment.

Also in the present study, *Dipylidium sp* was another parasite found to be highly prevalent. This parasite is one of the most common tapeworms infesting companion animals (Rousseau *et al.* 2022). It is a zoonotic parasite responsible for a disease named dipylidiasis worldwide (Benitez-Bolivar *et al.* 2022). Humans acquire infection through fecal-oral transmission by incidental ingestion of infected intermediate host (fleas) containing the cycstercoid of *Dipylidium sp* (Rousseau *et al.* 2022). Young children who closely interact with dogs, such as kissing them or being licked by infected pet dogs, are at a higher risk of acquiring infections (García-Agudo *et al.* 2014).

## CONCLUSION

In conclusion, the presence of these parasites in stray dog populations within the study areas highlights their contribution to environmental contamination and their potential impact on public health. The prevalence of intestinal parasitic infections in animal and human populations has been linked to the degree of environmental contamination with parasite eggs or oocysts. Therefore, it is essential to implement coordinated efforts to reduce stray dog roaming in Suleja metropolis, and by extension, across Niger State, Nigeria.

## REFERENCES

Abbas, I., Baghdadi, H.B., Rizk, M.A., El-Alfy, E.S., Elmishmishy, B. and Gwida, M. (2023). Gastrointestinal parasites of dogs in Egypt: An update on the prevalence in Dakahlia governorate and a meta-analysis for the published data from the Country. *Animals (Basel)*, 13(3): 496. <https://doi.org/10.3390/ani13030496>

Adesoye, O.A., Adediran, A.D., Oyeniyi, T.O., Olagundoye, E.O., Izekor, R.T., Adetunji, O.O., Babalola, A.S., Adeniyi K., Akinsete, I., Oyeniran, O., Akinleye, C., Isaac, C. Adekeye, T. and Adeogun, A. (2024). Implication of Larval Breeding Sites on Diversity of Mosquito Species in Suleja Metropolis, Northcentral Nigeria. *Dutse Journal of Pure and Applied Sciences*, 10 (1): 401-410.

Akeredolu, A.B. and Sowemimo, O.A. (2014). Prevalence, intensity and associated risk factors for *T. canis* infection in Nigerian dogs. *Journal of Parasitology and Vector Biology*, 6(8): 111-116.

Archer, C. E., Appleton, C. C., Mukaratirwa, S., Lamb, J. and Corrie Schoeman, M. (2017). Endo-parasites of public-health importance recovered from rodents in the Durban metropolitan area, South Africa. *Southern African Journal of Infectious Diseases*, 32(2): 57–66. <https://doi.org/10.1080/23120053.2016.1262579>

Ayinmode, A.B., Obebe, O.O. and Olayemi, I. (2016) Prevalence of potentially zoonotic gastrointestinal parasites in canine feces in Ibadan, Nigeria. *Ghana Medical Journal*, 50(4): 201-206.

Benitez-Bolivar, P., Rondón, S., Ortiz, M., Díaz-Díaz, J., León, C. and Riveros, J. (2022). Morphological and molecular characterization of the parasite *Dipylidium caninum* infecting an infant in Colombia: A case report. *Parasites Vectors*, 15(1): 463; <https://doi.org/10.1186/s13071-022-05573-4>.

Bhattarai, G.P. (2020). Gastrointestinal helminth parasites in stray and pet dogs of Kathmandu valley, Nepal. (*Doctoral dissertation, Department of Zoology*). 275-300

Cimino, R.O., Jeun, R., Juarez, M. Pamela S., P., Vargas, Adriana, E., Patricia, E., Julio N., Alejandro K. and Rojelio

M. (2015). Identification of human intestinal parasites affecting an asymptomatic peri-urban Argentinian population using multi-parallel quantitative real-time polymerase chain reaction. *Parasites Vectors*, 8: 380. <https://doi.org/10.1186/s13071-015-0994-z>

Del Brutto, O.H., Rajshekhar, V., White Jr, A.C., Tsang, V.C., Nash, T.E. and Takayanagui O.M. (2018). Proposed diagnostic criteria for neurocysticercosis. *Neurology*, 90(15): 738-746.

Duncan, K.T., Koons, N.R., Litherland, M.A., Little, S.E. and Nagamori, Y. (2020). Prevalence of intestinal parasites in faecal samples and estimation of parasite contamination from dog parks in central Oklahoma. *Veterinary Parasitology: Reg St.*, 19: 100362.

Ezema, K.U., Malgwi, S.A., Zango, M.K., Kyari, F., Tukur, S.M. and Mohammed, A. (2019). Gastrointestinal parasites of dogs (*Canis familiaris*) in Maiduguri, Borno State, Northeastern Nigeria: Risk factors and zoonotic implications for human health. *Veterinary World*, 12(7): 1150-1153.

García-Agudo, L., García-Martos, P. and Rodríguez-Iglesias, M. (2014). *Dipylidium caninum* infection in an infant: A rare case report and literature review. *Asian Pacific Journal of Tropical Biomedicine.*, 4: 565-567.

Issa, A.R., Mero, W.M., Arif, S.H. and Casulli, A. (2022). Prevalence of Taeniid eggs in the feces of stray dogs collected from different location of Zakho city, Kurdistan Region, Iraq. *Academic journal of Nawroz University*, 11(4): 259-265.

Jajere, S.M., Lawal, J.R., Shittu, A., Waziri, I., Goni, D.M., Fasina, F.O. (2022). Epidemiological study of gastrointestinal helminths among dogs from Northeastern Nigeria: a potential public health concern. *Parasitology Research*, 121(7): 2179-2186.

Kamani, J., Massetti, L., Olubade, T., Balami, J.A., Samdi, K.M. and Traub, R.J. (2021). Canine gastrointestinal parasites as a potential source of zoonotic infections in Nigeria: A nationwide survey. *Preventive Veterinary Medicine*, 192:105385; <https://doi.org/10.1016/j.prevetmed.2021.105385>

Kebede, N. (2019). Prevalence of gastrointestinal parasites of dogs and community awareness about zoonotic diseases in Chagni town, northwestern Ethiopia. *Ethiopian Veterinary Journal*, 23(2): 13-26.

Kirman, R., Akyuz, M., Balkaya, İ., Güven, E. and Avcioglu, H. (2023). Gastrointestinal helminths of stray dogs in Erzurum province: Prevalence and risk to public health. *Ankara Üniversitesi Veteriner Fakültesi Dergisi*, 70(3): 345- 348.

Lyons, M.A., Malhotra, R. and Thompson, C.W. (2022). Investigating the free-roaming dog population and gastrointestinal parasite diversity in Tulum, México. *PLoS One*, 17(10): e0276880.

- Magaji, A.A., Mohammed, M.N., Saulawa, M.A. and Salihu, M.D. (2012) Survey of zoonotic gastrointestinal parasites of dogs (*Canis familiaris*) slaughtered at Zuru area, Kebbi state, Nigeria. *Journal of Advanced Veterinary Research*, 1(5): 132-136.
- Matthew, T.T., Seer, I.J. and David, O.K. (2016). The prevalence of gastrointestinal helminths (GIH) infection of dogs in Makurdi metropolis. *Imperial Journal of Interdisciplinary Research*, 2(8): 1042-1049.
- Mizgajska-Wiktor, H., Jarosz, W. and Fogt-Wyrwas, R. (2017). *Toxocara* spp. infection and its negative associations with childhood asthma in urban settings. *Parasitology Research*, 116(2): 515-520.
- Moro, K.K., Abah, A.E. (2019). Epizootiology of zoonotic parasites of dogs in Abua area of Rivers State, Nigeria. *Veterinary and Animal Science Journal*, 7: 100045; <https://doi.org/10.1016/j.vas.2018.100045>
- Odeniran, P.O. and Ademola, I.O. (2013). Prevalence of zoonotic gastrointestinal helminth in dogs and knowledge of the risk of infection by dog owners in Ibadan, Nigeria. *Nigerian Veterinary Journal*, 34(3): 212-221
- Omonijo, A.O. and Mukaratirwa, S. (2023). Knowledge and practices on consumption of free-range chickens in selected rural communities of KwaZulu-Natal, South Africa, with focus on zoonotic transmission of *Toxoplasma gondii* and *Toxocara* spp. *Tropical Animal Health and Production*, 55(1): 9.
- Phoosangwalthong, P., Luong, N.H., Wongwigkan, J., Kamyinkird, K., Phasuk, J. and Pattanatanang, K. (2022). *Toxocara canis* and *Toxocara cati* in stray dogs and cats in Bangkok, Thailand: Molecular prevalence and risk factors. *Parasitology*, 2(2): 88-94.
- Rousseau J., Castro, A., Novo, T. and Maia, C. (2022). *Dipylidium caninum* in the twenty-first century: epidemiological studies and reported cases in companion animals and humans. *Parasites Vectors*. 15: 131. <https://doi.org/10.1186/s13071-022-05243-5>
- Rubinsky-Elefant, G., Hirata, C.E., Yamamoto, J.H. and Ferreira, M.U. (2010). Human toxocarasis: Diagnosis, worldwide seroprevalences and clinical expression of the systemic and ocular forms. *Annals of tropical medicine and parasitology - NCBI*, 104(1): 3-23.
- Souza, C.T.V.D., Dorr, A.P., Silva, V.L.D.B., Silva, F.D.L., Silva, E.B.D. and Ramos, D.G.D.S. (2023). Occurrence of gastrointestinal parasites in dogs from Cuiabá, Mato Grosso. *Revista Brasileira de Parasitologia Veterinária*, 32: e012422.
- Sowemimo, O.A. (2009). The prevalence and intensity of gastrointestinal parasites of dogs in Ile-Ife, Nigeria. *J. Helminthol.*, 83(1): 27-31.
- Stafford, K., Kollasch, T.M., Duncan, K.T., Horr, S., Goddu, T. And Heinz-Loomer, C. (2020). Detection of gastrointestinal parasitism at recreational canine sites in the USA: the DOGPAPCS study. *Parasites Vectors*, 13(1): 1-10.
- Sukupayo, P.R. and Tamang, S. (2023). Prevalence of zoonotic gastrointestinal helminth parasite among dogs in Suryabinayak, *Veterinary Medicine International*, 12: 234 <https://doi.org/10.1155/2023/3624593>.
- Suliaman, Y., Zakaria, M.A. and Pengsakul, T. (2020). Prevalence of intestinal helminth parasites of stray dogs in Shendi area, Sudan. *Annals of Parasitology*, 66(1): 115-118.
- Ugbomoiko, U.S., Ariza, L. and Heukelbach, J. (2008). Parasites of importance for human health in Nigerian dogs: high prevalence and limited knowledge of pet owners. *BMC Veterinary Research*, 4(1): 1-9.
- Ünal, G.G. and Gokpinar, S. (2022). Prevalence of intestinal parasites in dogs and its importance in terms of public health. *International Journal of Veterinary and Animal Research*, 2020; 3(3): 64-68.
- Wu, Q., Richard, M., Rutschmann, A. Miles, D. and Clobert, J. (2019). Environmental variation mediates the prevalence and co-occurrence of parasites in the common lizard, *Zootoca vivipara*. *BMC Ecology*, 19: 44. <https://doi.org/10.1186/s12898-019-0259-3>.