

Sahel Journal of Life Sciences FUDMA (SAJOLS) June 2025 Vol. 3(2): 205-210 ISSN: 3027-0456 (Print) ISSN: 1595-5915 (Online) DOI: <u>https://doi.org/10.33003/sajols-2025-0302-25</u>



**Research Article** 

# Transmission Dynamics of Virulent Newcastle Disease Virus (vNDV) in Chickens Experimentally Vaccinated with Newcastle Disease Virus Lasota and Challenged with vNDV

\*Tasiu Mallam Hamisu<sup>1</sup>, Halilu Abdullahi<sup>1</sup>, Yasheruram Muhammad Shettima<sup>1</sup>, Hauwa Yauba<sup>1</sup> and Sani Mohammed<sup>2</sup>

<sup>1</sup>Department of Veterinary Microbiology, Faculty of Veterinary Medicine, University of Maiduguri, Nigeria

<sup>2</sup>Department of Veterinary Public Health and Preventive Medicine, Faculty of Veterinary Medicine, University of Maiduguri, Nigeria

\*Corresponding Author's email: <u>tasiugln@unimaid.edu.ng</u>; Phone: +2348068064657

## ABSTRACT

Newcastle Disease (ND), caused by Newcastle Disease Virus (NDV), poses a significant threat to the global poultry industry, affecting thousands of avian species and causing substantial economic losses. Despite the use of vaccines such as La Sota to induce immunity, sterilizing immunity that completely prevents virus shedding has not been achieved. This study evaluated NDV shedding and body weight changes in chickens experimentally inoculated with virulent NDV (vNDV), with or without prior vaccination, and assessed transmission to contact (sentinel) birds. Cloacal swab samples were collected at different time points post-challenge (1, 7, 14, and 21 days) and analysed using the haemagglutination (HA) test. Results revealed that in unvaccinated, vNDV-inoculated chickens, the highest virus shedding occurred at day 7, followed by a significant decline at days 14 and 21. In contrast, vaccinated-challenged chickens exhibited highest virus shedding at 24 hours post-challenge, with titres decreasing over time, and no virus shedding was detected in their contact birds. Across both studies, contact birds consistently showed higher body weight gains compared to inoculated or vaccinated-challenged birds. These findings highlight the effectiveness of the La Sota vaccine in reducing virus shedding and limiting transmission, though further research is needed to elucidate the molecular mechanisms underlying NDV shedding and immunity.

# Keywords: Chickens; HA; La Sota; NDV; vNDV Shedding

**Citation:** Hamisu, T.M., Abdullahi, H., Shettima, Y.M., Yauba, H. & Mohammed, S. (2025). Transmission Dynamics of Virulent Newcastle Disease Virus (Vndv) in Chickens Experimentally Vaccinated with Newcastle Disease Virus Lasota and Challenged with vNDV. *Sahel Journal of Life Sciences FUDMA*, 3(2): 205-210. DOI: <u>https://doi.org/10.33003/sajols-2025-030222-25</u>

# INTRODUCTION

Newcastle disease (ND) is an acute, highly contagious viral disease that affects a variety of avian hosts, including poultry and non-poultry species. It has a wide range of clinical and pathological symptoms as well as morbidity and mortality rates, including neurological, respiratory, and gastrointestinal symptoms (Miller *et al.*, 2013). The age of the birds, their immunological health, and the virulence of the Newcastle Disease virus

(NDV) strain affect how severe ND is (Alexander, 2000). The virus remains a significant threat to poultry despite widespread use of vaccines like La Sota Newcastle disease virus shedding is of epidemiological significant toward the spread of ND, this is because the spread of NDV begins with the shedding of the virus by the infected/ carrier bird and the subsequent inhalation of contaminated aerosol by susceptible bird (Hamisu *et al.*, 2022).

Studies have highlighted critical challenges in vaccine efficacy and transmission dynamics, particularly when vaccinated chickens are exposed to virulent NDV (vNDV) strains (Habibi et al., 2014; Ferreira et al., 2020; Mahmood et al., 2023; Hanan, et al., 2025). While conventional La Sota vaccines reduce clinical signs and mortality, they often fail to prevent infection, viral replication, and shedding of heterologous genotypes like the prevalent genotype VII (Van Boven et al., 2008; Bwala et al., 2012). This mismatch allows vNDV to persist and transmit even in vaccinated flocks, undermining control efforts (Van Boven et al., 2008; Wajid et al., 2018; Sedeik et al., 2019). Experimental evidence demonstrates that La Sota-vaccinated chickens challenged with genotype VII vNDV exhibit substantial viral shedding through respiratory and cloacal routes, facilitating transmission to susceptible birds (Van Boven et al., 2008; Bwalaet al., 2012). Notably, a study found that vaccination via the ocular route provided better protection against tracheal virus shedding compared to cloacal administration (Bwalaet al., 2012, Palya et al., 2021). However, even birds with low antibody titres, though protected from severe disease, can sustain viral transmission chains, with reproduction numbers (R) exceeding 1 in partially immune populations (Van Boven et al., 2008; Bwalaet al., 2012).

Advances in recombinant vaccines, such as rLa Sota-HN expressing genotype VII haemagglutininneuraminidase (HN) protein, show promise in addressing these gaps. Trials indicate enhanced protection, reduced shedding, and lower transmission rates compared to classical La Sota (Bu *et al.*, 2019).

Several studies have shown that the amount of NDV shed determines whether or not susceptible birds will be infected (Costa-Hurtado *et al.*, 2015; Ferreira *et al.*, 2019). Whether the titre of the vNDV shed can cause infection to susceptible birds in a La Sota vaccinated-challenged setting remains generally unknown. This underscores the need to evaluate the La Sota vaccination strategy to account for both individual bird protection and herd immunity thresholds.

# MATERIALS AND METHODS

# Study Area and Experimental Design

This study was conducted in Maiduguri, Borno State, Nigeria, characterized by a semi-arid climate with distinct rainy and dry seasons.

A total of 12-day-old chicks were purchased from Sayeed Breeders, Jos, Plateau State, Nigeria, and raised at the Animal Facility of the Veterinary Teaching Hospital, University of Maiduguri, with feed and water provided *ad libitum*. The chickens were broadly divided into two groups of 6 chickens each, challenge group and vaccinated-challenge group.

For the challenge group: At three weeks old, three chicks were inoculated with 0.1 ml of  $10^{6}$ ELD<sub>50</sub>vNDV, while three served as non-inoculated contact birds.

For the vaccinated-challenge experiment: At two weeks old, three chicks were vaccinated with 1 ml La Sota vaccine and challenged with 0.1 ml of 10<sup>6</sup>ELD<sub>50</sub>vNDVat six days post-vaccination, while the other three chicks served as contact birds.

## Sample Collection and Processing

Cloacal swab samples were collected from all the birds at 1, 7, 14, and 21 days post-challenge, placed in viral transport media, and transported in a cold chain to Virology laboratory, Department of Veterinary Microbiology, Faculty of Veterinary Medicine, University of Maiduguri, where they were stored at -20°C before the samples were subjected to the haemagglutination (HA) test.

## Haemagglutination Test

The HA test was conducted following a standard protocol (Hierholzer *et al.,* 1969; Kaufmann *et* 

*al.*, 2017). Briefly, chicken red blood cells (RBCs) were prepared by washing the blood collected from NDV-free chickens in Phosphate Buffered Saline (PBS) and making a 1% RBC suspension. Thereafter, 25  $\mu$ L of PBS was dispensed into each well of the V-bottom microtiter plates. A 25  $\mu$ L of the test sample was then added to the plate, and two-fold serial dilution was performed across the plate using a multichannel pipette. Then, 25  $\mu$ L of PBS was further added to each well. Finally, 25  $\mu$ L of 1% chicken RBC was added to each well. The sides of the plate were gently tapped to ensure proper mixing. The plate was allowed to stand for 45 minutes at room temperature.

#### Data Analysis

Data generated were presented in tables. Difference in means between the groups was analyzed using one-way Analysis of Variance in SPSS-IBM, USA (v25.0). Values of P<0.05 were considered statistically significant throughout the study.

# RESULTS

#### Virus Shedding Dynamics

In unvaccinated, vNDV-inoculated chickens, the highest vNDV shedding was observed at day 7 postchallenge, with a significant decline by days 14 and 21. The difference in shedding between day 7 and other days was statistically significant (p<0.05). In contact birds, only a slight, non-significant increase in shedding was noted at days 14 and 21 (Table 1). In La Sota vaccinated-vNDV challenged chickens, the highest vNDV shedding occurred at 24 hours post-challenge, with titres decreasing at subsequent time points (7, 14, and 21 days). There was a statistically significant difference between day 1 shedding and later time points (p<0.05). No virus shedding was detected in contact birds throughout the study (Table 2).

#### **Body Weight Gain**

Contact birds consistently showed higher average body weight gains at six weeks of age compared to

their inoculated or vaccinated-challenged counterparts. Specifically, contact birds gained 3.0– 3.5 kg, with contact birds in the vaccinated challenge group having highest body weight, while inoculated or vaccinated-challenged birds gained 2.83–3.13 kg, with the vNDV inoculated group having the lowest body weight (Tables 3 and 4).

Table 1: Means of vNDV shedding at Various Time Points in Chickens Inoculated with vNDV and Contact Birds				
Sampling Pariod (days)	Maan VNDV Titra + SD (85% CI)			

Sampling Period (days)	wean vivov Titre ± SD (95%CI)	
	Challenged birds	Contact birds
1	1: 0.00±0.00 <sup>a</sup>	1:0.0±0.00 <sup>a</sup>
7	1:53.33 <b>±</b> 22.03 <sup>b</sup>	1: 0.0 ±0.00 <sup>a</sup>
14	1:2.00±0.00ª	1: 0.667 <b>±</b> 1.15ª
21	1:8.00±0.00 <sup>a</sup>	1: 0.667 <b>±</b> 1.15ª

Values with the same letters are not statistically different between sampling periods within a treatment group. Values with different letters differed significantly within the sampling periods in a treatment group. The value is the mean ± SEM of three samples

Table 2: Mean NDV Titre at Various Time Points in Chickens Vaccinated with NDV La Sota and Challenged with	I
vNDV	_

Sampling Period (days)	Mean vNDV ⊺itre ± SD (95%CI)		
	Vaccinated- challenge birds	Contact birds	
1	1:13.3 ± 4.62ª	0.00 <sup>a</sup>	
7	$1:0 \pm 0.00^{b}$	0.00 <sup>a</sup>	
14	$1:0 \pm 0.00^{b}$	0.00 <sup>a</sup>	
21	1:6 ± 3.46 <sup>b</sup>	0.00ª	

Values with the same letters are not statistically different between sampling periods within a treatment group. Values with different letters differed significantly within the sampling periods in a treatment group. The value is the mean ± SEM of three samples

Groups	Body Weight (Kg)			Average
Challenge birds	2.7	2.9	2.9	2.83
Contact birds	3.0	3.0	3.0	3.0

Table 4: Body Weight Gain of Six-Week-Old Vaccinated-Challenged and Contact Birds						
Groups	Body	v Weight (Kg)		Average		
Vaccinated-Challenge birds	3.1	3.1	3.2	3.13		
Contact birds	3.5	3.5	3.5	3.5		

#### DISCUSSION

Newcastle Disease remains a persistent challenge for poultry health due to the high mortality rates associated with virulent NDV strains and the economic impact of the disease outbreaks (Moustapha *et al.*, 2023; NIp*et al.*, 2024). In this study, we evaluated the impact of La Sota vaccination in relation to vNDV challenge, and assessed whether the titre of the vNDV shed in both vaccinated-challenge and vNDV inoculated group only, can cause infection to susceptible birds.

In unvaccinated chickens challenged with vNDV, the highest viral shedding occurs at day 7 post-

infection, followed by a marked decline by days 14 and 21, with statistically significant differences between day 7 and days14 and 21 (p< 0.05). This pattern aligns with studies showing higher cloacal shedding in naturally exposed, unvaccinated village chickens, which exhibit a 43.8% shedding prevalence (Sajo *et al.*, 2023). The delayed peak may reflect initial viral replication before immune activation, while the subsequent decline suggests partial innate immune control or resource depletion in hosts (Mark *et al.*, 2013). Contact birds in these groups show only minor, non-significant increases in shedding at later stages, likely due to environmental contamination or staggered transmission dynamics (Dimitrov, 2024).

In contrast, La Sota-vaccinated chickens display rapid virological control, with peak shedding at 24 hours post-challenge followed by significant reductions at days 7–21 (p< 0.05). This accelerated response correlates with enhanced intraepithelial lymphocyte (IEL) populations, including CD3<sup>+</sup>, CD4<sup>+</sup>, and CD8<sup>+</sup> cells, which are higher in vaccinated birds compared to unvaccinated controls (Baftiet al., 2020; Hamisu *et al.*, 2022). However, in contact birds, no vNDV has been recorded. The absence of shedding in contact birds from vaccinated groups demonstrates sterilizing immunity, likely mediated by mucosal IgA and systemic IgG responses that block both clinical disease and transmission (Baftiet *al.*, 2020; Amer *et al.*, 2023).

Contact birds consistently achieved higher average body weight gains (3.0–3.5 kg at six weeks) compared to both vNDV-inoculated and La Sotavaccinated-challenged birds (2.83–3.13 kg), with the lowest weights in the vNDV-inoculated group. Among the contact birds, those in the vaccinatedchallenge environment had the highest body weights.

Infection with vNDV leads to severe clinical signs, including gastrointestinal haemorrhages, dehydration, emaciation, and necrotic lesions, all of which contribute to poor feed intake and nutrient absorption, resulting in reduced body weight gain (Moustapha *et al.*, 2023).

The observed higher body weight gains in contact birds compared to vNDV-inoculated or La Sotavaccinated groups may be due to the facts that contact birds likely experienced subclinical or milder infections due to controlled viral exposure, minimizing metabolic disruptions. Studies show that NDV infection reduces feed efficiency and weight gain by impairing pancreatic function and nutrient absorption (Wang et al., 2015; Abdienet al., 2025). Furthermore, direct inoculation introduces a higher viral load, triggering pronounced immune responses that divert energy from growth (Ellakanyet al., 2018). For example, vNDV-inoculated birds in one study had a 37.5% mortality rate and prolonged viral shedding (9 days), exacerbating weight loss (Ellakanyet al., 2018).

La Sota vaccination, while protective, induces transient stress that can suppress growth. Research demonstrates that repeated NDV vaccinations reduce early-stage body weight gain (1–21 days) due to corticosterone release and metabolic adaptation (Wang *et al.*, 2015). It also aligns with the findings that vaccinated broilers often exhibit lower feed conversion ratios (FCR) than unvaccinated controls (Bakhtiar *et al.*, 2022). A 2022

study comparing La Sota and Mukteswar vaccines found unvaccinated controls outperformed vaccinated groups in weight gain, underscoring the trade-off between immune protection and growth (Bakhtiar *et al.*, 2022).

In summary, contact birds are not exposed to acute immune activation and metabolic stress associated with direct inoculation or vaccination, enabling superior weight gain.

# CONCLUSION

This study reveals distinct patterns in viral shedding and growth outcomes between unvaccinated and La Sota-vaccinated chickens challenged with vNDV. In unvaccinated, inoculated birds, viral shedding peaked sharply at day 7 post-challenge, followed by a significant decline by days 14 and 21 (p < 0.05), while contact birds in this group showed only minimal, non-significant shedding at later stages. In contrast, La Sota-vaccinated challenged chickens exhibited rapid but short-lived shedding, with titres highest at 24 hours post-challenge before dropping markedly by day 7, and no shedding was detected in their contact counterparts. Notably, contact birds across both groups consistently achieved superior weight gains (3.0-3.5 kg) compared to directly exposed birds (2.83-3.13 kg), with the lowest growth observed in vNDV-inoculated chickens. These findings demonstrate that La Sota vaccination not only curtails viral shedding intensity and duration but also reduces transmission to contacts, while unvaccinated infection imposes measurable growth lost. The enhanced performance of contact birds further highlights the indirect benefits of limiting viral exposure, reinforcing vaccination as a critical strategy for both disease control and productivity optimization in poultry industry.

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

#### **AUTHORS CONTRIBUTIONS**

TMH and YMS designed the research, draft and reviewed the manuscript; HA and HY conducted the experiment; SM analayzed the data.

#### REFERENCES

Abdien, H. M. F., El-Dimerdash, M. M. Z., Morsy, E. A., Ali, N. M. and Abdallah, M. S. (2025). Comparative Analysis of Commercial Recombinant Vaccination Strategies Against Newcastle Disease in Broilers.*German Journal of Veterinary Research*, *5*(1): 17–29.

https://doi.org/10.51585/gjvr.2025.1.0116

 Alexander, D. J. (2000). Newcastle Disease and

 Other Avian Paramyxoviruses. *Rev. Sci. Tech. Off. Int. Epiz.*, 19(2): 443–462.

 https://doi.org/10.1002/9781119421481.ch3

Amer, S. A. M. A., Ghetas, A. M., Maatouq, A. M., Ahmed, H. M., El-Bayoumi, K. M., Bosila, M. A. E. R. and El-Shemy, A. A. (2023). Enhanced Protection and Blocked Viral Shedding for Broiler Chickens in Challenge with Newcastle Disease Virus Genotype VII by Generation of Oil Inactivated Vaccine Antigenically-Matched to The Endemic Virus in Egypt. Advances in Animal and Veterinary Sciences, 11(9): 1548–1556.

https://doi.org/10.17582/journal.aavs/2023/11.9. 1548.1556

Bafti, S., Mosleh andDadras, H. (2020). Assessment of Type I Interferons, Clinical Signs and Virus Shedding in Broiler Chickens with Pre and Post Challenge Newcastle Disease Vaccination. *Iran Journal of Veterinary Research*, 21(2):84-91. PMID: 32849885; PMCID: PMC7430373.

Bakhtiar, A., Mehmood, S. A., Bhatti, A. R., Ahmad, S., Khalid, N., Igbal, J., Nadeem, A and Ahmad, W. (2022). Comparative Efficacy of Newcastle Disease's Live Vaccines in Broilers using Hemagglutination Inhibition (HI) Test at JabaMansehra. Pakistan Journal of Agricultural Research, 35(4): 629-636. https://doi.org/10.17582/JOURNAL.PJAR/2022/35. 4.629.636

Bu, Y. wen, Yang, H. ming, Jin, J. hui, Zhao, J., Xue, J and Zhang, G. zhong. (2019). Recombinant Newcastle Disease Virus (NDV) La Sota Expressing the Haemagglutinin–Neuraminidase Protein of Genotype VII NDV Shows Improved Protection Efficacy Against NDV Challenge. *Avian Pathology*, *48*(2): 91–97.

https://doi.org/10.1080/03079457.2018.1548754 Bwala, D. G., Clift, S., Duncan, N. M., Bisschop, S. P. R. andOludayo, F. F. (2012). Determination of the Distribution of Lentogenic Vaccine and Virulent Newcastle Disease Virus Antigen in the Oviduct of SPF and Commercial Hen Using Immunohistochemistry. Research in Veterinary Science. 93(1): 520-528. https://doi.org/10.1016/j.rvsc.2011.06.023

Dimitrov, K. (2024). Newcastle Disease in Poultry (Avian Pneumoencephalitis, Exotic Newcastle Disease).

https://www.msdvetmanual.com/poultry/newcast le-disease-and-other-p...

Ellakany, H. F., Gado, A. R., Elbestawy, A. R., Abd El-Hamid, H. S., Hafez, H. M., Abd El-Hack, M. E., Swelum, A. A., Al-Owaimer, A. and Saadeldin, I. M. (2018). Interaction Between avian Influenza Subtype H9N2 and Newcastle Disease Virus Vaccine Strain (Lasota) in Chickens. *BMC Veterinary*  Research, 14(1). <u>https://doi.org/10.1186/s12917-</u> 018-1689-4

https://doi.org/10.1016/j.vaccine.2020.06.004. Habibi, H., Nili, H., Asasi, K., Mosleh, N., Firouzi, S., and Mohammadi, M. (2014). Efficacy and transmissibility of Newcastle disease I-2 vaccine strain against a field isolate of virulent ND virus (JF820294.1) in village chicken. *Tropical Animal Health and Production*, 47, 73 - 78. https://doi.org/10.1007/s11250-014-0687-1.

Hanan M. F. Abdien, Mohsen M. Z. El-Dimerdash, Eslam A. Morsy, Norhan M. Ali, and Mona S. Abdallah (2025). Comparative analysis of commercial recombinant vaccination strategies against Newcastle disease in broilers. *German Journal of Veterinary Research*. https://doi.org/10.51585/gjvr.2025.1.0116.

Hamisu, T. M., Aliyu, H. B., Hair-Bejo, M., Omar, A. R. and Ideris, A. (2022). Alteration in the Population of Intraepithelial Lymphocytes and Virus Shedding in Specific-Pathogen-Free Chickens Following Inoculation with Lentogenic and Velogenic Newcastle Disease Virus Strains. *Viral Immunology*, *35*(4):328–337.

https://doi.org/10.1089/vim.2021.0148

Hierholzer, J. C., Suggs, M. T. and Hall, E. C. (1969).StandardizedViralHemagglutinationandHemagglutinationInhibitionTests.II.DescriptionandStatisticalEvaluation.Appliedmicrobiology,18(5):824-833.

https://doi.org/10.1128/am.18.5.824-833.1969 Kaufmann, L., Syedbasha, M., Vogt, D., Hollenstein, Y.,Hartmann, J., Linnik, J. E. and Egli, A. (2017). An Optimized Hemagglutination Inhibition (HI) Assay to Quantify Influenza-Specific Antibody Titers. *Journal of Visualized Experiments*, (130), e55833. https://doi.org/10.3791/55833

Mahmood, S., Skinner, P., Warren, C., Mayers, J., James, J., Núñez, A., Lean, F., Brookes, S., Brown, I., Banyard, A., and Ross, C. (2023). In vivo challenge studies on vaccinated chickens indicate a virus genotype mismatched vaccine still offers significant protection against NDV.. Vaccine. https://doi.org/10.1016/j.vaccine.2023.12.037.

Mark D., J., Christopher, J. W., Jeanne, M. F. and Jannifer, C. O. (2013). Birds Shed RNA-Viruses According to the Pareto Principle. *PLOS ONE*, *8*(8): e72611.

Miller, P. J., Afonso, C. L., El Attrache, J., Dorsey, K. M., Courtney, S. C., Guo, Z. and Kapczynski, D. R. (2013). Effects of Newcastle Disease Virus Vaccine Antibodies on the Shedding and Transmission of Challenge Viruses. *Developmental and Comparative Immunology*, 41(4): 505–513. https://doi.org/10.1016/j.dci.2013.06.007

Moustapha, A., Talaki, E., Akourki, A. and Ousseini, M. (2023). Newcastle Disease Virus in Poultry: Current Status and Control Prospects. *World's Veterinary Journal*, *13*(2): 240–249. https://doi.org/10.54203/scil.2023.wvj26

Palya, V., Tatár-Kis, T., Arafa, A., Felföldi, B., Mató, T., and Setta, A. (2021). Efficacy of a Turkey Herpesvirus Vectored Newcastle Disease Vaccine against Genotype VII.1.1 Virus: Challenge Route Affects Shedding Pattern. *Vaccines*, 9. https://doi.org/10.3390/vaccines9010037.

Rasoli, M., Yeap, S. K., Tan, S. W., Moeini, H., Ideris, A., Bejo, M. H., Alitheen, N. B. M., Kaiser, P., and Omar, A. R. (2014). Alteration in Lymphocyte Responses, Cytokine and Chemokine Profiles in Chickens Infected with Genotype VII and VIII VelogenicNewcastle Disease Virus. *Comparative Immunology, Microbiology and Infectious Diseases*, *37*(1): 11–21.

https://doi.org/10.1016/j.cimid.2013.10.003

Sajo, M. U., Hamisu, T. M., Saidu, Haruna, N. M., Shettima, El-Yuguda, Abubakar, M. B., Madu and Waziri, M. M. (2023). Comparative Analysis of Newcastle Disease Virus Shedding from Naturally Infected Breeds of Poultry in Maiduguri, Nigeria. Sahel Journal of Veterinary Science20(3): 33–37. https://doi.org/10.54058/saheljvs.v%vi%i.380

Sedeik, M., Elbestawy, A., El-Shall, N., El-Hack, M., Saadeldin, I., Saadeldin, I., Swelum, A., and Swelum, A. (2019). Comparative efficacy of commercial inactivated Newcastle disease virus vaccines against Newcastle disease virus genotype VII in broiler chickens. *Poultry Science*, 98, 2000–2007. https://doi.org/10.3382/ps/pey559.

Van Boven, M., Bouma, A., Fabri, T. H. F., Katsma, E., Hartog, L. and Koch, G. (2008). Herd immunity to Newcastle Disease Virus in Poultry by Vaccination. *Avian Pathology*, *37*(1): 1–5. https://doi.org/10.1080/03079450701772391

Wajid, A., Basharat, A., Bibi, T., and Rehmani, S. (2018). Comparison of protection and viral shedding following vaccination with Newcastle disease virus strains of different genotypes used in vaccine formulation. *Tropical Animal Health and Production*, 50, 1645 - 1651. https://doi.org/10.1007/s11250-018-1607-6.

Wang, X., Zhou, Q., Shen, J., Yao, J. and Yang, X. (2015). Effect of Difference Doses of Newcastle Disease Vaccine Immunization on Growth Performance, Plasma Variables and Immune Response of Broilers. *Journal of Animal Science and Biotechnology*, 6(1).

https://doi.org/10.1186/s40104-015-0019-y.