



Research Article

Prevalence of Haemozoin among Patients with *Plasmodium falciparum* Malaria in Some Selected Hospitals in Bakori and Funtua, Katsina State, Nigeria

A. M. Umar¹, M. Sani¹, E. J. Joshua¹ and A. Ado²

¹Department of Biological Sciences, Faculty of Life Sciences, Federal University Dutsin-Ma, P.M.B. 5001 Dutsin-Ma Katsina State, Nigeria

²Department of Microbiology, Faculty of Life Sciences, Federal University Dutsin-Ma, P.M.B. 5001 Dutsin-Ma Katsina State, Nigeria

*Corresponding Author's email: sanimuhammadamb1089@gmail.com

ABSTRACT

In malaria diagnosis and accurate assessment are critical to clinical management. Intraleukocytic malaria pigment characteristics were explored as it relates to severe falciparum malarial disease. This study evaluates the prevalence of haemozoin among patients with plasmodium falciparum malaria in some selected hospitals in Bakori and Funtua, Katsina state, Nigeria. Intraleukocytic malaria pigment (hemozoin) were detected on thin films by counting 500 leukocytes and determining the proportions of haemozoin-containing neutrophils, lymphocytes and monocytes. A total of 392 blood samples were stained and examined under the light microscope using 100x objective lens. Immunochromatography tests based on the capture of the parasite antigen from the peripheral blood using monoclonal anti-HRP-II antibodies were used for detecting Plasmodium falciparum antigens, as well as the presence and quantity of the malaria pigment in leukocytes in smears. The findings of this study recorded the highest number of intraleukocytic malaria pigment within age group 2-10years, with 40.8% prevalence rate collectively, and the prevalence of haemozoin related to sex, the females had a high prevalence compare to males this could be due to a combination of biological factors, social inequalities and challenges in accessing preventive care.

Keywords: Hemozoin; Immunochromatography; Intraleukocytic; Leukocytes; Neutrophis

Citation: Umar, A.M., Sani, M., & Joshua, E.J. (2026). Prevalence of Haemozoin among Patients with *Plasmodium falciparum* Malaria in Some Selected Hospitals in Bakori and Funtua, Katsina State, Nigeria. *Sahel Journal of Life Sciences FUDMA*, 4(1): 398-403. DOI: <https://doi.org/10.33003/sajols-2026-0401-47>

INTRODUCTION

Malaria is one of the most common and important parasitic diseases worldwide. It affects the lives of almost all people living in the area of Africa defined by the southern Sahara Desert in the north, and latitude of about 28° in the south (WHO/UNICEF, 2003). About 40% of the world's population lives in malaria-endemic areas. Over 90% of all cases of life-threatening malaria occur among African children. Majority of deaths occur in children under five years of age. Severe malaria is the commonest cause of death, particularly in rural areas that are not serviced by formal health systems. Malaria-associated immunosuppression has been widely investigated. It

has been suggested that malaria pigment, also known as haemozoin may participate in the mechanisms underlying this immunosuppression (Coban *et al.*, 2002). For prognosis, malaria is a potentially life-threatening parasitic disease caused by infection with *Plasmodium* protozoa transmitted by an infective female *Anopheles* mosquito. *Plasmodium falciparum* infection carries a poor prognosis with a high mortality if untreated, but it has an excellent prognosis if diagnosed early and treated appropriately.

Rapid and accurate diagnosis and prognostic assessment are critical to clinical management. In 6027 prospectively studied patients diagnosed with

severe malaria we assess the prognostic value of peripheral blood film counts of malaria pigment containing polymorphonuclear leukocytes (PMNs) and monocytes have been assessed among patients with severe malaria (White, 2022). Currently malaria control is heavily dependent on chemotherapy, to which resistance is quickly evolving in endemic regions (Nosten *et al.*, 2000). One of the main interventions of the global malaria control strategy for effective disease management is prompt and accurate diagnosis (WHO, 1993). In the tropics, malaria diagnosis is carried out mainly by microscopic methods while rapid diagnostic techniques (RDTs) are available mainly in non-endemic settings (Fitri *et al.*, 2022). This study aimed to evaluate the prevalence of haemozoin among patients with *Plasmodium falciparum* malaria in some selected hospitals in Bakori and Funtua, Katsina state, Nigeria.

MATERIALS AND METHODS

Study Area

A randomized study was conducted in Comprehensive Hospital Bakori and General Hospital

Funtua (Figure 1). The study was carried-out over a period of five months from February 2024 to June 2024.

Immunochromatography Test (ICT) tests based on the capture of the parasite antigen from the peripheral blood using monoclonal anti-HRP-II antibodies and anti-aldolase antibodies against the parasite antigen targets were used (Moody, 2002).

Thick and thin blood films as described by Benishangul (2002), were made on clean slides and labelled accordingly as recommended by (WHO, 2002). To prepare for each thick film, 2 drops of blood sample were laced on a clean grease-free glass by means of a pasteur pipette and, the blood will be gently spread for 20 seconds using the corner of a second slide to defibrinate the blood and to obtain a round smear. The slide was immersed for 30 minutes in the staining trough, containing Giemsa solution prepared with buffered water in the ratio of 1:20. Thereafter the slide was rinsed and allowed to dry (Benishangul, 2002).

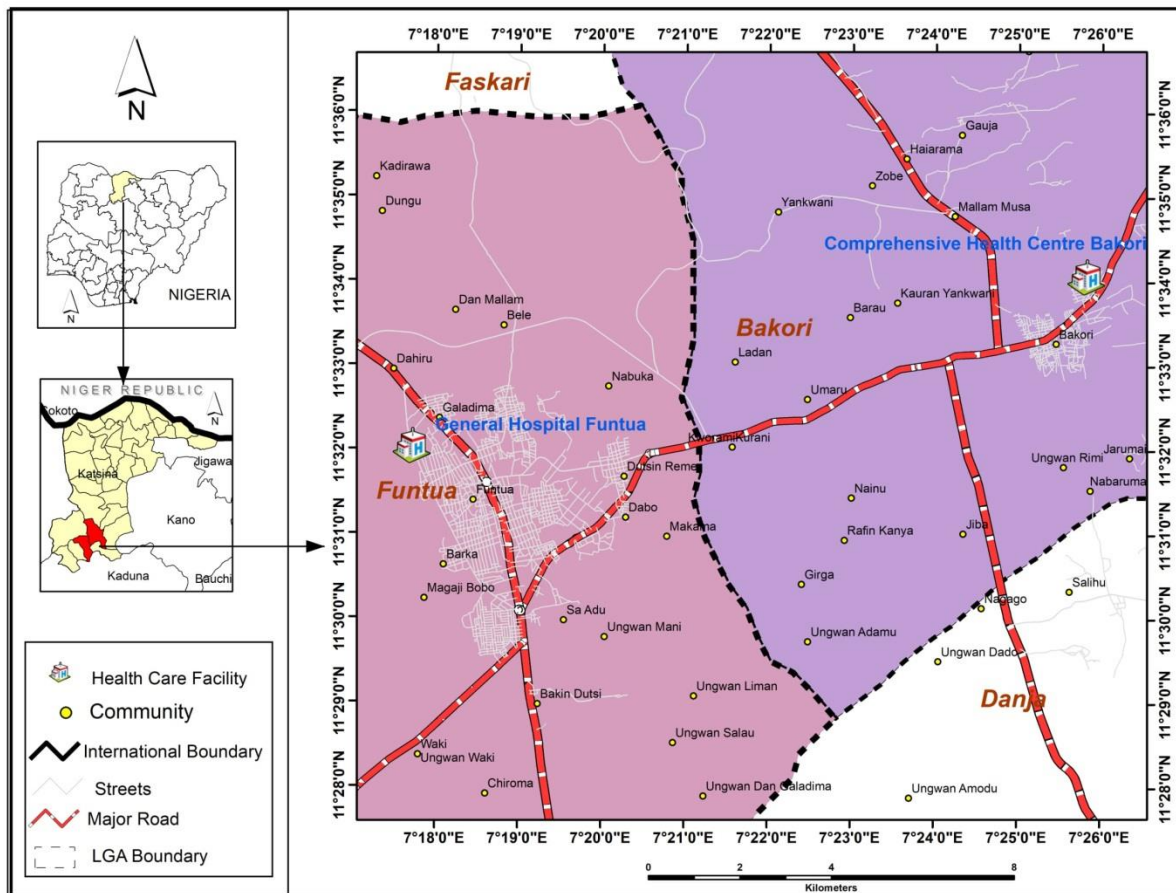


Figure1 map of Bakori and Funtua showing sampled Hospitals (GIS FUDMA 2025)

This were done using thin smears. The thin smears were fixed in absolute methanol for one minute and before staining. These smears were then stained using Giemsa. Then white blood cell differential counts manually determined. Intraleukocytic malaria pigment were detected on thin films by counting 500 leukocytes and determining the proportions of haemozoin-containing neutrophils, lymphocytes and monocytes.

Microsoft Excel and Statistical Package for Social Science (SPSS Version 20 IBM Statistics) software was used to analyse data. Inferential statistics where data was presented in the results as tables and figures. Chi square was used for prevalence association between gender and age groups based on disease severity.

RESULTS

A total of 392 were examined, 264 (67%) had a mild malaria and 59 (15.1%) had a severe malaria. The chi-square ($\chi^2 = 784$) and p-value = 0.000 shows that there was significant association between

parasitaemia with pigment (Table 1) Therefore the null (H_0) is rejected, which states there is no present of the intraleukocytic malaria pigment (haemozoin) in a patient with severe *falciparum* malaria.

General hospital Funtua had a higher prevalence of 51.5% when compared to comprehensive hospital Bakori which had a prevalence of 48.5%. The chi-square ($\chi^2 = 3.588$) and p-value = 0.166 shows that there is no significant association between parasitaemia and health facility (Table 2).

The prevalence of parasitaemia in relation to sex, the females had a high prevalence of 55.4% compared to males 44.6%. The chi-square ($\chi^2 = 1.149$) and p-value = 0.563 shows that there is no significant association between parasitaemia and sex (Table 3).

The age group 2-10yrs had the highest prevalence of 40.8% malaria, while the age group 71-80yrs had the lowest prevalence of 0.3% malaria and the chi-square ($\chi^2 = 14.268$) and p-value = 0.579 shows that there is no significant association between parasitaemia and age group (Table 4).

Table 1: The prevalence of parasitaemia in respect to pigment in Bakori and Funtua (n=392)

Parasitaemia/Pigment level	Frequency	Prevalence (%)
Mild (P+)	264	67.3
Moderate (P++)	69	17.6
Severe (P+++)	59	15.1
χ^2	784.000	
p-value	0.000	

Table 2: The prevalence of parasitamaia in relation to health facility

Parasitaemia	Health facility	
	CHB	GHF
P+	120	144
P++	40	29
P+++	30	29
Total	190	202
Prevalence (%)	48.5%	51.5%
χ^2		3.588
p-value		0.166

CHB: Comprehensive Hospital Bakori. GHF: General Hospital Funtua

Table 3: The prevalence of parasitaemia in relation to sex

Parasitaemia	Sex		Total
	Male	Female	
P+	114	150	
P++	31	38	
P+++	30	29	
Total	175	217	392
Prevalence (%)	44.6	55.4	
χ^2			1.149
p-value			0.563

Table 4: The prevalence of parasitaemia in relation to age group

Parasitaemia	Age group									Total
	1m-1yr	2-10yrs	11-20yrs	21-30yrs	31-40yrs	41-50yrs	51-60yrs	61-70yrs	71-80yrs	
P+	46	110	31	27	19	12	10	9	0	
P++	10	27	6	8	7	8	2	0	1	
P+++	9	23	7	6	6	4	3	1	0	
Total	65	160	44	41	32	24	15	10	1	392
Prevalence (%)	16.6	40.8	11.2	10.5	8.2	6.1	3.8	2.6	0.3	
χ^2										14.268
p-value										0.579

DISCUSSION

The prevalence 15.1% of severe malaria was recorded in the study localities. This prevalence is lower than 43.1% reported by Wogu *et al.* (2017) in Rivers State, Nigeria. General hospital Funtua had a higher prevalence of 51.5% when compared to comprehensive hospital Bakori which had a prevalence of 48.5%. The high prevalence in Funtua could be due to the presence of some water canals from Mai Ruwa reservoir and Gwagwaye reservoir that passes around the town of Funtua and other clogged gutters and drains. The prevalence of parasitaemia in relation to sex, the females had a high prevalence compare to males this could be due to a combination of biological factors, social inequalities and challenges in accessing preventive care. This finding is not in line with the findings of Raasti *et al.*, (2024). The findings of this study recorded the highest number of intraleukocytic malaria pigment within age group 2-10 had 40.8 %. This prevalence could be due to low immunity at that age group. This finding is not in line with the findings of Raasti *et al.*, (2024) who recorded high prevalence in age group of 40-60 years. This study is in line with findings (Nwaorgu, and Orajaka, 2011) in Awka North Local Government Area, Anambra State South East Nigeria.

CONCLUSION

This study recorded prevalence of severe malaria an overall 15.1% within the hospitals in Bakori (CHC) and (GHF) Funtua. Most of severe malarial cases based on my research were associated to female due to presence haemozoin and some accessed biological factors. Intraleukocytic malaria pigment (aemozoin) was more prevalent among 2-10years. More research should be done using Sysmex XN450 Haematology Analyzer Full Blood Count which can

also provide information on the viability of complete blood count.

Microscopic examination proved to be more effective in detection of low parasite density; as such, microscopic examination should be constantly and widely employed in laboratory for adequate diagnosis and detection of malaria parasite and its kind. Parasitaemia is important in diagnosis of malaria, it should not be used only as an indicator of malaria severity.

Governmental and non-governmental organization should be continued carry out awareness campaign on the use of ITNs, Environmental hygiene, as well as preventive drugs to reduce the incidence in the study area. Also, proper environmental sanitation is required to remove mosquitoes from their breeding.

REFERENCES

Adams, J. H., Sim, B. K., & Dolan, S. A. (1992). A family of erythrocyte binding proteins of malaria parasites. *Proceedings of the National Academy of Sciences of the USA*, 89(1), 7085–7089.

Alano, P. (2007). *Plasmodium falciparum* gametocytes: Still many secrets of a hidden life. *Molecular Microbiology*, 66(2), 291–302.

Alles, H. K., Mendis, K. N., & Carter, R. (1998). Malaria mortality rates in South Asia and in Africa: *Implications for malaria control. Parasitology Today*, 14(9), 369–375.

Bannister, L. H., Hopkins, J. M., Fowler, R. E., Krishna, S., & Mitchell, G. H. (2001). A brief illustrated guide to the ultrastructure of *Plasmodium falciparum* asexual blood stages. *Parasitology Today*, 16(10), 427–433.

Baum, J., Maier, A. G., Good, R. T., Simpson, K. M., & Cowman, A. F. (2005). Invasion by *P. falciparum*

- merozoites suggests a hierarchy of molecular interactions. *PLoS Pathogens*, 1(3), e37.
- Billker, O., Lindo, V., Panico, M., (1998). Identification of xanthurenic acid as the putative inducer of malaria development in the mosquito. *Nature*, 392(6673), 289–292.
- Carter, R., & Mendis, K. N. (2002). Evolutionary and historical aspects of the burden of malaria. *Clinical Microbiology Reviews*, 15(4), 564–594.
- Cavalier-Smith, T. (2003). Protist phylogeny and the high-level classification of protozoa. *European Journal of Protistology*, 39(4), 338–348.
- Chitnis, C. E., & Sharma, A. (2008). Targeting the *Plasmodium vivax* Duffy-binding protein. *Trends in Parasitology*, 24(1), 29–34.
- Conroy, A., McDonald, C., & Kain, K. (2012). Malaria in pregnancy: Diagnosing infection and identifying foetal risk. *Expert Review of Anti-infective Therapy*, 10(11), 1331–1342.
- Cowman, A. F., & Crabb, B. S. (2006). Invasion of red blood cells by malaria parasites. *Cell*, 124(4), 755–766.
- Cox-Singh, J., & Singh, B. (2008). Knowlesi malaria: Newly emergent and of public health importance. *Trends in Parasitology*, 24(9), 406–410.
- Cyrklaff, M., Kudryashev, M., Leis, A., et al. (2007). Cryoelectron tomography reveals periodic material at the inner side of subpellicular microtubules in apicomplexan parasites. *Journal of Experimental Medicine*, 204(6), 1281–1287.
- Djoufounna, J., Mayi, M. P. A., & Bamou, R. (2022). High prevalence of asymptomatic *Plasmodium falciparum* malaria in Madenene, a locality in the forest savannah transition zone Centre Region of Cameroon. *Current Research in Parasitology & Vector-Borne Diseases*, 2, Article 1104.
- Frevert, U., Engelmann, S., Zougbedeb, S., (2005). Intravital observation of *Plasmodium berghei* sporozoite infection of the liver. *PLoS Biology*, 3(6), 1034–1046.
- Gardner, M. J., Hall, N., Fung, E., White, O., Berriman, M., Hyman, R. W., Carlton, J. M., Pain, A., Nelson, K. E., Bowman, S., Paulsen, I. T., James, K., Eisen, J. A., Fraser, C. M., & Barrell, B. (2002). Genome sequence of the human malaria parasite *Plasmodium falciparum*. *Nature*, 419(6906), 498–511.
- Goldberg, D. E. (2005). Hemoglobin degradation. *Current Topics in Microbiology and Immunology*, 295, 275–291.
- Kasetsirikul, S., Buraanpong, J., Srituravanich, W., Kaewthamason, M., & Pimpin, A. (2016). The development of malaria diagnostic techniques: A review of the approaches with focus on dielectrophoretic and magnetophoretic methods. *Malaria Journal*, 15(1), Article 358.
- Khan, S. M. (2005). Proteome analysis of separated male and female gametocytes reveals novel sex-specific *Plasmodium* biology. *Cell*, 121(5), 675–687.
- Krampa, F. D., Aniweh, Y., Awandare, G. A., & Kanyong, P. (2017). Recent progress in the development of diagnostic tests for malaria. *Diagnostics*, 7(3), Article 54.
- Ladda, R., Aikawa, M., & Sprinz, H. (1969). Penetration of erythrocytes by merozoites of mammalian and avian malarial parasites. *Journal of Parasitology*, 55(3), 470–478.
- Lyke, K. E., Diallo, D. A., & Dicko, A. (2003). Association of interleukocytic *Plasmodium falciparum* malaria pigment with disease severity, clinical manifestation, and prognosis in severe malaria. *Clinical Infectious Diseases*, 33(5), 39–40.
- Mauritz, J. M. A., Esposito, A., & Ginsburg, H. (2009). The homeostasis of *Plasmodium falciparum*-infected red blood cells. *PLoS Computational Biology*, 5(4), e1000339.
- Mayfong, M., Pukrittayakamee, S., Newton, P. N., & White, N. J. (2004). Mixed species malaria infections in humans. *Trends in Parasitology*, 20(5), 233–240.
- Mendis, K., Sina, L. J., Marchesini, P., & Carter, R. (2001). The neglected burden of *Plasmodium vivax* malaria. *American Journal of Tropical Medicine and Hygiene*, 64(12), 97–106.
- Miller, L. H., Mason, S. J., Dvorak, J. A., McGinniss, M. H., & Rothman, I. K. (1975). Erythrocyte receptors for (*Plasmodium knowlesi*) malaria: *Duffy blood group determinants*. *Science*, 189(4202), 561–563.
- Miller, S. A., & Marley, J. P. (1999). *Zoology* (4th ed.). WMC Brown Publishers.
- Murray, C. J. L., & Lopez, A. D. (1997). Mortality by cause for eight regions of the world: Global burden of disease study. *The Lancet*, 349(9061), 1269–1276.
- Murray, C. K., & Bennett, J. W. (2009). Rapid diagnosis of malaria. *Interdisciplinary Perspectives on Infectious Diseases*, 2009, Article 415953.
- Nwaorgu, O. C., & Orajaka, B. N. (2011). Prevalence of malaria among children 1–10 years old in communities in Awka North Local Government Area, Anambra State South East Nigeria. *African Journal of Science, Technology and Society*, 22(5), 1–8.
- Okello, P. E., Van Bortel, W., Byaruhanga, A. M., Correwyn, A., Roelants, P., Talisuna, A., D'Alessandro, U., & Coosemans, M. (2006). Variation in malaria transmission intensity in seven sites throughout Uganda. *American Journal of Tropical Medicine and Hygiene*, 75(2), 219–225.

- Olszewski, K. L., Morrisey, J. M., Wilinski, D., et al. (2009). Host–parasite interactions revealed by *Plasmodium falciparum* metabolomics. *Cell Host & Microbe*, 5(2), 191–199.
- Omoya, F. O., & Ajayi, K. O. (2020). Prevalence of malaria among febrile patients attending government hospitals in Ondo State, South-West Nigeria. *American Journal of Epidemiology and Public Health*, 4(4), 17–24.
- Pei, X., Guo, X., & Coppel, R. (2007). The ring-infected erythrocyte surface antigen (RESA) of *Plasmodium falciparum* stabilizes, 110(3), 1036-1042.
- Prusty, S. K. R., & Das, B. S. (2001). Low incidence of the severe complications of malaria and absence of malaria-specific mortality, in Tensa, Sundergarh district, Orissa State, India, an area hyperendemic for malaria. *Annals of Tropical Medicine & Parasitology*, 95(2), 133–140.
- Raasti, A., Nasir, O., Khalid, M. A., Zafar, F., Khan, W., & Nadeem, S. F. (2024). Evaluating malaria prevalence across different age and gender groups in Peshawar through light microscopic analysis. *Journal of Health and Habitation Research*, 4(2), 767–771.
- Rowe, A., & Kyes, S. A. (2004). The role of *Plasmodium falciparum* var genes in malaria in pregnancy. *Molecular Microbiology*, 53(4), 1011–1019.
- Snow, R. W. (1997). Relation between severe malaria morbidity in children and level of *Plasmodium falciparum* transmission in Africa. *The Lancet*, 349(9066), 1650–1654.
- Weiss, D. J., Lucas, T. C. D., Nguyen, M., Nandi, A. K., Bisanzio, D., Battle, K. E., Cameron, E., Twohig, K. A., Pfeffer, D. A., Rozier, J. A., & Gething, P. W. (2019). Mapping the global prevalence, incidence, and mortality of *Plasmodium falciparum*: A spatial and temporal modelling study. *The Lancet*, 394(10195), 322–331.
- Weiss, D. J., Lucas, T. C. D., Nguyen, M., Nandi, A. K., Bisanzio, D., Battle, K. E., Cameron, E., Twohig, K. A., Pfeffer, D. A., Rozier, J. A., Gibson, H. S., Rao, P. C., Casey, D., Bertozzi-Villa, A., Collins, E. L., Dalrymple, U., Gray, N., Harris, J. R., Howes, R. E., & Gething, P. W. (2019). Mapping the global prevalence, incidence, and mortality of *Plasmodium falciparum*, 2000–17: A spatial and temporal modelling study. *The Lancet*, 394(10195), 322–331.
- White, N. J. (2022). Leukocyte counts in falciparum malaria in African children from an endemic area. *PubMed*. 24(2):145–9
- WHO. (2023). World malaria report 2023. World Health Organization.
- Wogu, M. N., Nduka, F. O., & Wariso, K. T. (2017). Prevalence of uncomplicated and severe malaria in outpatients of a tertiary hospital in Rivers State, Nigeria. *Journal of Applied Life Sciences International*, 15(3), 1–5.
- World Health Organization. (2000). The world health report 2000: Making a difference. World Health Organization.
- World Health Organization. (2017). Report on infectious diseases: Scaling up the response to infectious diseases (pp. 13–16).
- World Health Organization. (2018). World malaria report 2018.
- World Health Organization. (2022). World malaria report 2022: Tracking progress and gaps in the global response to malaria. Retrieved February 16, 2023, from [URL]
- Yeoh, S., O'Donnell, R. A., Koussis, K., (2007). Subcellular discharge of a serine protease mediates release of invasive malaria parasites from host erythrocytes. *Cell*, 131(6), 1072–1083.
- Yu, M., Kumar, T. R. S., Nkrumah, L. J., (2008). The fatty acid biosynthesis enzyme FabI plays a key role in the development of liver-stage malarial parasites. *Cell Host & Microbe*, 4(6), 567–578.